

Source: Dr. D. Vijaya Ramu, BT, VU

16BT404 GENOMICS AND PROTEOMICS

Hours Per Week :

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3	-	2	4	

Course Description and Objectives:

This course offers various tools available for analyzing genomes and proteomes of different organisms. The objective of this course is to enrich students with a wide array of genomic and proteomic approaches utilized for various therapeutic applications.

Course outcomes:

Upon completion of the course, the student will be able to

- CO1. Identify exons and introns in a given genomic sequence to understand organization of genomes.
- CO2. Subclone and express genes in prokaryotic expression vector and detect proteins in polyacrylamide gels.
- CO3. Gain adequate knowledge on various tools available for annotation of genomes.
- CO4. Apply concepts of Microarrays.
- CO5. Comprehnd techniques of protein-separation, sequencing and identification.

SKILLS:

- ✓ Amplify selectable marker.
- Design primers for amplification of screenable marker in construct.
- Identify exons and introns in Th1 genes of different species.
- ✓ Identify cSNPs between any two genes pertaining to two different breeds of chick.
- ✓ Expression of genes using prokaryotic systems.

Genomics and Proteomics

UNIT - 1

INTRODUCTION TO GENOMICS: Organization and structure of genomes; Genome size; Sequence complexity; Introns and exons; Chromosomes-isolation, chromosome micro dissection and its applications.

UNIT - 2

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ACTIVITIES:

o Amplification of

• Expertise in

designing

primers.

• Analyzing

exons and

o Expertise with

o Expertise with

expression of

gene.

gene.

introns of gene.

sub-cloning of

gene by PCR.

GENE IDENTIFICATION AND EXPRESSION: Genome annotation; Traditional routes of gene identification; Detecting open-reading frames; Software programs for finding genes; Identifying the function of a new gene; Gene ontology; Overview of comparative genomics; Determining gene function by sequence comparison and through conserved protein structure; Global expression profiling; Traditional approaches to expression profiling.

UNIT - 3

ANALYSIS OF PROTEOMICS: Introduction to Proteomics - the proteome, mining proteomes, bridging genomics and proteomics data; Analysis of proteomes - SDS-PAGE, 2D gel electrophoresis, detecting proteins; Mass spectrometry.

UNIT - 4

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ANALYSIS OF GENOMICS: Micro arrays - types of micro arrays, designing a microarray experiment, applications of microarray technology; Chip array; Shotgun method.

UNIT - 5

APPLICATIONS OF GENOMICS AND PROTEOMICS: Insights from genome sequencing of various species - human, mouse, *Plasmodium falciparum*, *Saccharomyces cerevisiae* and *Mycobacterium tuberculosis*; Application of proteome analysis in drug development and toxicology; Applications of proteomics in plant genetics and breeding.

LABORATORY EXPERIMENTS

LIST OF EXPERIMENTS

- 1. Insilico determination of exons and introns in a gene.
- 2. Designing of primers employing generunner software.
- 3. Amplification of gene by polymerase chain reaction.
- 4. Sequence analysis of gene by BLAST.
- 5. Insilico identification of cSNPs.
- 6. Cloning of gene in plasmid for gene modifications.
- 7. Expression of gene for functional protein production.
- 8. Analyzing the solubility of proteins by plasmolysis.

TEXT BOOKS:

- 1. S. B. Primrose and R.M. Twyman, "Principles of Genome Analysis and Genomics", 7th edition, Blackwell Publishing, 2006.
- 2. S. Sahai, "Genomics and Proteomics, Functional and Computational Aspects" Plenum Publication, 1999.

REFERENCE BOOKS:

- 1. A. K. Konopka and J. C. Crabbe, "Compact Hand Book Computational Biology", Marcel Dekker, USA, 2004.
- 2. S.R. Pennington and M.J. Dunn, "Proteomics: From Protein Sequence to Function", 1st edition, Academic Press, San Diego, 1996.

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Total hours 30

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